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**REVIEW** 

# The Isoform Spesific Roles of Rho-Kinases in Vascular Diseases

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### **ABSTRACT**

A small GTPase RhoA and its downstream effectors Rho-associated protein kinases (ROCK) signaling pathway activation mediate smooth muscle contraction. ROCKs inhibit myosin light chain phosphatase (MLCP) dephosphorylation and therefore reduce relaxation. However, nitric oxide (NO) that is produced and released from endothelial cells has an inhibitory effect on the ROCK pathway in vasculature. Studies in which ROCK activity was inhibited by variety of pharmacological agents (HA1077 or Y-27632) have shown that it has some critical effects on systemic diseases like hypertension or diabetes mellitus. Indeed this activity may show isoform specificity (ROCK1 or ROCK2) dependent on the pathology. Therefore, in vascular pathogenesis ROCK pathway with its isoforms also need to be considered due to its direct effects on the vasoconstriction.

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**Key words:** Rho Kinases; ROCK1; ROCK2; Vasoconstriction; Vascular diseases

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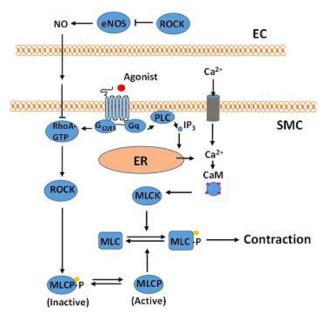
### INTRODUCTION

Rho-associated protein kinases (ROCKs) play a critical role in smooth muscle contraction and relaxation. After activation of small GTPase RhoA its effector protein ROCK mediates contraction. Briefly after a signal arrives to cell membrane and activates membrane receptors or voltage operated Ca<sup>2+</sup> channels (VOCC) free cytoplasmic Ca<sup>2+</sup> concentration increases. Then Ca<sup>2+</sup> binds to calmodulin and activates myosin light chain kinase (MLCK). MLCK phosphorylates myosin light chains (MLC), which are the regulatory subunits of the myosin heads.

MLCK phosphorylates (MLC) the subunits of the myosin heads. Phosphorylated MLCs enable the cross bridges between myosin and actin and so contraction occurs in smooth muscles[1,2]. On the other hand RhoA mediated ROCK is the other mediator of contraction with Ca<sup>2+[3]</sup>. Once activated ROCK provides continuation of contractile activity by inhibiting MLC phosphatase (MLCP) which dephosphorylate MLC and induces relaxation in smooth muscle cells. Together with Ca<sup>2+</sup>, ROCK pathways precisely control the vasoconstriction. In the arteries smooth muscle contraction is directly effects blood pressure by regulating the vessel diameter and tension<sup>[1,4]</sup>. In addition to vasoconstrictor effectors, endothelial derived Nitric Oxide (NO) is a vasodilator agent for smooth muscle cells which regulates the relaxation through cGMP pathway and also reduces ROCK activity and thereby contraction<sup>[5-8]</sup> (Figure 1). In the regulation of the vascular tonus these vasoconstrictor and vasodilator pathways mediate contraction. In recent years many studies has shown that Rho Kinase pathway should be taken into consideration in treatments of vascular diseases[9,10]

ROCK consists of two isoforms ROCK1 and ROCK2. ROCK1 enzyme is expressed in a plenty of different tissues like lung, kidney, stomach whereas ROCK2 is mostly expressed in heart, brain and skeletal muscle<sup>[11]</sup>. Cellular localization of the ROCK1 and ROCK2 also show diversity. ROCK1 is mostly localized at plasma membrane but ROCK2 at centrosomes of smooth muscle cells. At cardiomyocytes ROCK2 localized at intercalated discs, and at skeletal muscle cells Z-discs and sarcoplasmic reticulum<sup>[12]</sup>.

Although, they have high genetic homology in their kinase domain



**Table 1** The contribution of ROCK pathway onto the contraction of smooth muscle. ROCK enhance contraction by phosphorylating MLCP. This inactive form of MLCP could not dephosphorylate MLC which leads to attenuation of the muscle relaxation. ROCK activity is reduced with the NO through cGMP pathway. Abbreviations: PLC: Phospholipase C, IP3: Inositol (1,4,5)-trisphosphate, eNOS: Endothelial nitric oxide synthase, MLC: Myosin light chains, MLCK: MLC kinase, MLCP: MLC phosphatase, ROCK: Rho kinase, NO: Nitric oxide.

(92%)<sup>[13]</sup>, Yoneda *et al*<sup>[14]</sup> (2005) showed that they have isoform specific functions even in the same cell. According to this study ROCK1 is more active in focal adhesion and fiber formations rather than ROCK2 in primary rat embryo fibroblasts. Conversely, ROCK2 is the primer player in phagocytic activity. In another study it was shown that ROCK1 and ROCK2 have distinct roles in adhesion and differentiation in keratinocytes<sup>[15]</sup>. Also the experiments performed with the cells which derived from ROCK1 and 2 knockout animals, was shown that ROCK 1 acts in MLC2 phosphorylation and cell detachment, whereas ROCK2 in actin cytoskeleton stabilization<sup>[16]</sup>. Their mechanistic difference was indicated by Wang Y *et al*<sup>[17]</sup> (2009) that ROCK2 can bind directly to the myosin-binding subunit (MYPT1) of myosin phosphatase but not ROCK1. This difference reflects that ROCK1 use intermediate components for MLCP inhibition and this fringed pathway selection makes harder to understand the ROCK activity.

However lack of the isoform specific ROCK blockers (HA1077 and Y27632 are nonspecific blockers of ROCK) make difficult to distinguish the functional roles of the isoforms. But such a critical pathway that regulates constrictive mechanisms in vascular system deserves more precise evaluation. Therefore in this review we tried to we focus on these functional differences between two isoforms in vasculature from diseases perspective.

# ROCK ISOFORMS IN ARTERIAL AND PUL-MONARY ARTERIAL HYPERTENSION

Smooth muscle cells primarily regulate vascular volume and thereby blood pressure in the aorta. The role of the vasoconstrictors on the hypertension is a detailed examined in many studies concluded ROCK also play a major role in persistency of the high pressure in aorta<sup>[2,18]</sup>

It was shown in the hypertensive animal models that ROCK medi-

ated vasoconstriction is involved arterial hypertension by blocking its function with Y-27632<sup>[19]</sup>. Other selective ROCK inhibitor HA1077 named as fasudil is believed to be a key therapeutic for human use. In one study dealing with hypertensive patients it was shown that the fasudil induce a vasodilator effect on the arterial pressure<sup>[20]</sup>. Also in a study Fukumoto *et al*<sup>[21]</sup> showed the effects of the fasudil on the patients with pulmonary arterial (PA) hypertension. The treatment with fasudil hydrochloride caused a slight decrease in the PA hypertension.

In both of these arterial high pressure diseases differences in the expression levels of ROCK1 and ROCK2 were observed. The immunostaining experiments ROCK2 (but not ROCK1) showed that its expression increases in arterioles of the lung sections taken from the PA hypertension patients<sup>[22]</sup>. The same study indicated that the hypoxia induced PA hypertension with vascular smooth muscle specific ROCK2 gene knockout mice, the right ventricular systolic pressure was significantly reduced versus control. Their findings indicate the importance of ROCK2 for the development of hypoxia-induced PA hypertension. Also ROCK2 gene silencing was improved erectile function on spontaneously hypertensive rats suggesting ROCK2 inhibition can be used as a specific therapeutic target for vascular dysfunctions caused by hypertension<sup>[23]</sup>.

### **ATHEROSCLEROSIS**

When dealing with this very complicated inflammatory disease we see that on tunica intima, the layer surrounded with the formations such foam cells (monocytes/macrophages) that decrease vessel diameter and even make it more stiffening<sup>[24,25]</sup>. Impaired endothelium activity (endothelium dysfunction) causes dysregulation of NO release, which was thought as a major responsible factor for the initiation of atherosclerosis<sup>[26,27]</sup>. According to the study of Anju Nohria *et al*<sup>[28]</sup> ROCK inhibition with fasudil caused endothelial dependent vasodilation in the patients with coronary artery disease. Their measurements with brachial artery ultrasonography suggest the relation between endothelium activity and ROCK inhibition in atherosclerosis. In another study with mice ROCK inhibition with Y27632 results a protection against atherosclerosis by reducing significantly size of the atherosclerotic plaque formation significantly<sup>[29]</sup>.

The individual roles of ROCK1 and ROCK2 in atherosclerosis tried to be explained in several studies. ROCK1 knockout was decreased atherosclerotic lesion formations in aortas from the bone marrow (BM) derived macrophage transplanted LDLr knockout mice<sup>[30]</sup>. While the experiments with ROCK2 lacking in the cultured BM differentiated macrophages was shown the importance of ROCK2 in the foam cell formations<sup>[31]</sup>.

### **DIABETES**

Type independently, diabetes mellitus (DM) patients frequently suffer from the complications of circulatory system diseases such as cardiovascular or other vascular diseases. These complications may accompany with hypertension, atherosclerosis and thereby some ischemic diseases or systemic dysfunctions (peripheral, pulmonary, renin-angiotensin)<sup>[32-35]</sup>. It was shown that Rho kinase has a promoter effect on Ca<sup>2+</sup> sensitive vasocontraction with PKC in STZ induced DM model studies<sup>[36]</sup>. Also in the study by Sandu OA *et al*<sup>[37]</sup> (2001) the interaction of insulin with Rho kinase from phosphatidylinositol 3-kinase (PI3-kinase) and iNOS activated NO-cGMP pathway was specified in vascular smooth muscle cells (VSMC). According to them insulin receptor activation inhibits ROCK activity by

the NO pathway and a defectiveness in this pathway in diabetes and hypertension may lead an impaired relaxation with increased ROCK activity and resulting vasoconstriction. Also from the Rhokinase activity experiments it was observed that arteries from Zucker diabetic fatty (ZDF) rats or incubated with high glucose concentrations, ROCK activity increase parallel with the glucose concentration [38]. Rikiteka *et al* (2005) also has shown the correlation between vascular endothelial cells (HSVECs) and ROCK activity that increases in high glucose [39]. In the same study the high levels of Plasminogen activator inhibitor-1 (PAI-1) protein expression induced with hyperglycemia decreased in ROCK1 knockout (ROCK  $I^{+/-}$ ) murine lung endothelial cells. While PAI-1 is a risk factor in many vascular diseases [40], the effect of ROCK on the expression of this protein in hyperglycemia will also show the key role of ROCK activity in vascular dysfunctions.

In DM induced circulatory system diseases endothelial dysfunction which led impaired NO bioavailability causes impaired vasodilation. Many study show the effect of the ROCK pathway on the endothelial dysfunction and which then leads to impaired relaxation. In DM induced vascular endothelial dysfunction (VED) Rho kinase inhibition with fasudil improved eNOS/NO dependent vasodilation is stimulated by acetylcholine<sup>[41]</sup>. Also in diabetic retinopathy, a microvascular endothelial dysfunction, it was found that high glucose concentration has increased ROCK activity in retinal endothelial cell line, RF/6A cells<sup>[42]</sup>.

There are also ROCK1 and ROCK2 isoform specific studies in DM. Yao L (2013) by partly deletion both isoforms showed that ROCK1 is more effective in diabetic mice aorta according to vasorelaxive response to acetylcholine<sup>[43]</sup>. However in endothelial cells of rat thoracic aorta ROCK2 protein expression was found higher in DM with respect to the control group<sup>[44]</sup>. This difference may reflect different functional properties of ROCKs in the regulation of vascular smooth muscle contractions in DM.

Overall ROCK is a key player of many cellular functions. In recent years growing studies elicited its role in regulation of blood pressure in the vessels and therefore should be considered along with other contraction parameters. The isoforms ROCK1 and ROCK2 show branched functions, and regulate many diverse cellular activities on the circulatory system cells. Therefore, particularly in the treatment of cardiovascular diseases ROCKs with their isoforms should be taken into consideration because of their direct interventions on vasoconstriction

#### CONFLICT OF INTERESTS

There are no conflicts of interest with regard to the present study.

#### **REFERENCES**

- Webb, R. C. (2003) Smooth muscle contraction and relaxation. Advances in physiology education 27, 201-206
- Lee, D. L., Webb, R. C., and Jin, L. (2004) Hypertension and RhoA/Rho-kinase signaling in the vasculature: highlights from the recent literature. Hypertension 44, 796-799
- Berridge, M. J. (2008) Smooth muscle cell calcium activation mechanisms. The Journal of physiology 586, 5047-5061
- Zicha, J., Behuliak, M., Pinterova, M., Bencze, M., Kunes, J., and Vaneckova, I. (2014) The interaction of calcium entry and calcium sensitization in the control of vascular tone and blood pressure of normotensive and hypertensive rats. Physiological research / Academia Scientiarum Bohemoslovaca 63 Suppl 1, S19-27
- Michel, T., and Vanhoutte, P. M. (2010) Cellular signaling and NO production. Pflugers Archiv: European journal of physiology 459,

- 807-816
- Ignarro, L. J. (1989) Endothelium-derived nitric oxide: actions and properties. FASEB journal: official publication of the Federation of American Societies for Experimental Biology 3, 31-36
- Chitaley, K., and Webb, R. C. (2002) Nitric oxide induces dilation of rat aorta via inhibition of rho-kinase signaling. Hypertension 39, 438-442
- Wirth, A. (2010) Rho kinase and hypertension. Biochimica et biophysica acta 1802, 1276-1284
- Nunes, K. P., Rigsby, C. S., and Webb, R. C. (2010) RhoA/Rhokinase and vascular diseases: what is the link? Cellular and molecular life sciences: CMLS 67, 3823-3836
- Kataoka, C., Egashira, K., Inoue, S., Takemoto, M., Ni, W., Koyanagi, M., Kitamoto, S., Usui, M., Kaibuchi, K., Shimokawa, H., and Takeshita, A. (2002) Important role of Rho-kinase in the pathogenesis of cardiovascular inflammation and remodeling induced by long-term blockade of nitric oxide synthesis in rats. Hypertension 39, 245-250
- Nakagawa, O., Fujisawa, K., Ishizaki, T., Saito, Y., Nakao, K., and Narumiya, S. (1996) ROCK-I and ROCK-II, two isoforms of Rhoassociated coiled-coil forming protein serine/threonine kinase in mice. FEBS letters 392, 189-193
- Iizuka, M., Kimura, K., Wang, S., Kato, K., Amano, M., Kaibuchi, K., and Mizoguchi, A. (2012) Distinct distribution and localization of Rho-kinase in mouse epithelial, muscle and neural tissues. Cell structure and function 37, 155-175
- Noma, K., Oyama, N., and Liao, J. K. (2006) Physiological role of ROCKs in the cardiovascular system. American journal of physiology. Cell physiology 290, C661-668
- Yoneda, A., Multhaupt, H. A., and Couchman, J. R. (2005) The Rho kinases I and II regulate different aspects of myosin II activity. The Journal of cell biology 170, 443-453
- Lock, F. E., and Hotchin, N. A. (2009) Distinct roles for ROCK1 and ROCK2 in the regulation of keratinocyte differentiation. PloS one 4 e8190
- Shi, J., Wu, X., Surma, M., Vemula, S., Zhang, L., Yang, Y., Kapur, R., and Wei, L. (2013) Distinct roles for ROCK1 and ROCK2 in the regulation of cell detachment. Cell death & disease 4, e483
- Wang, Y., Zheng, X. R., Riddick, N., Bryden, M., Baur, W., Zhang, X., and Surks, H. K. (2009) ROCK isoform regulation of myosin phosphatase and contractility in vascular smooth muscle cells. Circulation research 104, 531-540
- Jalil, J., Lavandero, S., Chiong, M., and Ocaranza, M. P. (2005) [Rho/Rho kinase signal transduction pathway in cardiovascular disease and cardiovascular remodeling]. Revista espanola de cardiologia 58, 951-961
- Uehata, M., Ishizaki, T., Satoh, H., Ono, T., Kawahara, T., Morishita, T., Tamakawa, H., Yamagami, K., Inui, J., Maekawa, M., and Narumiya, S. (1997) Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. Nature 389, 990-994
- Masumoto, A., Hirooka, Y., Shimokawa, H., Hironaga, K., Seto-guchi, S., and Takeshita, A. (2001) Possible involvement of Rho-kinase in the pathogenesis of hypertension in humans. Hypertension 38, 1307-1310
- Fukumoto, Y., Matoba, T., Ito, A., Tanaka, H., Kishi, T., Hayashidani, S., Abe, K., Takeshita, A., and Shimokawa, H. (2005) Acute vasodilator effects of a Rho-kinase inhibitor, fasudil, in patients with severe pulmonary hypertension. Heart 91, 391-392
- Shimizu, T., Fukumoto, Y., Tanaka, S., Satoh, K., Ikeda, S., and Shimokawa, H. (2013) Crucial role of ROCK2 in vascular smooth muscle cells for hypoxia-induced pulmonary hypertension in mice. Arteriosclerosis, thrombosis, and vascular biology 33, 2780-2791
- Zhu, X., Lin, H., Jiang, R., Wang, R., Jiang, J., Peng, Q., and Fan, Z.
  (2014) Improving erectile function of spontaneously hypertensive rats by silencing ROCK2. Urology 84, 983 e911-988

- Bentzon, J. F., Otsuka, F., Virmani, R., and Falk, E. (2014) Mechanisms of plaque formation and rupture. Circulation research 114, 1852-1866
- Zieman, S. J., Melenovsky, V., and Kass, D. A. (2005) Mechanisms, pathophysiology, and therapy of arterial stiffness. Arteriosclerosis, thrombosis, and vascular biology 25, 932-943
- Davignon, J., and Ganz, P. (2004) Role of endothelial dysfunction in atherosclerosis. Circulation 109, III27-32
- Kratzer, A., Giral, H., and Landmesser, U. (2014) High-density lipoproteins as modulators of endothelial cell functions: alterations in patients with coronary artery disease. Cardiovascular research 103, 350-361
- Nohria, A., Grunert, M. E., Rikitake, Y., Noma, K., Prsic, A., Ganz, P., Liao, J. K., and Creager, M. A. (2006) Rho kinase inhibition improves endothelial function in human subjects with coronary artery disease. Circulation research 99, 1426-1432
- Mallat, Z., Gojova, A., Sauzeau, V., Brun, V., Silvestre, J. S., Esposito, B., Merval, R., Groux, H., Loirand, G., and Tedgui, A. (2003) Rho-associated protein kinase contributes to early atherosclerotic lesion formation in mice. Circulation research 93, 884-888
- Wang, H. W., Liu, P. Y., Oyama, N., Rikitake, Y., Kitamoto, S., Gitlin, J., Liao, J. K., and Boisvert, W. A. (2008) Deficiency of ROCK1 in bone marrow-derived cells protects against atherosclerosis in LDLR-/- mice. FASEB journal: official publication of the Federation of American Societies for Experimental Biology 22, 2561, 2570.
- Zhou, Q., Mei, Y., Shoji, T., Han, X., Kaminski, K., Oh, G. T., Ongusaha, P. P., Zhang, K., Schmitt, H., Moser, M., Bode, C., and Liao, J. K. (2012) Rho-associated coiled-coil-containing kinase 2 deficiency in bone marrow-derived cells leads to increased cholesterol efflux and decreased atherosclerosis. Circulation 126, 2236-2247
- Adler, A. I., Stratton, I. M., Neil, H. A., Yudkin, J. S., Matthews, D. R., Cull, C. A., Wright, A. D., Turner, R. C., and Holman, R. R. (2000) Association of systolic blood pressure with macrovascular and microvascular complications of type 2 diabetes (UKPDS 36): prospective observational study. Bmj 321, 412-419
- Moral-Sanz, J., Moreno, L., Cogolludo, A., and Perez-Vizcaino,
  F. (2014) Pulmonary vascular function in insulin resistance and diabetes. Current vascular pharmacology 12, 473-482
- Bugger, H., and Abel, E. D. (2014) Molecular mechanisms of diabetic cardiomyopathy. Diabetologia 57, 660-671
- 35. Jandeleit-Dahm, K., and Cooper, M. E. (2006) Hypertension and

- diabetes: role of the renin-angiotensin system. Endocrinology and metabolism clinics of North America 35, 469-490, vii
- Kizub, I. V., Pavlova, O. O., Johnson, C. D., Soloviev, A. I., and Zholos, A. V. (2010) Rho kinase and protein kinase C involvement in vascular smooth muscle myofilament calcium sensitization in arteries from diabetic rats. British journal of pharmacology 159, 1724-1731
- Sandu, O. A., Ito, M., and Begum, N. (2001) Selected contribution: insulin utilizes NO/cGMP pathway to activate myosin phosphatase via Rho inhibition in vascular smooth muscle. Journal of applied physiology 91, 1475-1482
- Bagi, Z., Feher, A., Cassuto, J., Akula, K., Labinskyy, N., Kaley, G., and Koller, A. (2011) Increased availability of angiotensin AT 1 receptors leads to sustained arterial constriction to angiotensin II in diabetes - role for Rho-kinase activation. British journal of pharmacology 163, 1059-1068
- Rikitake, Y., and Liao, J. K. (2005) Rho-kinase mediates hyperglycemia-induced plasminogen activator inhibitor-1 expression in vascular endothelial cells. Circulation 111, 3261-3268
- 40. Binder, B. R., Christ, G., Gruber, F., Grubic, N., Hufnagl, P., Krebs, M., Mihaly, J., and Prager, G. W. (2002) Plasminogen activator inhibitor 1: physiological and pathophysiological roles. News in physiological sciences: an international journal of physiology produced jointly by the International Union of Physiological Sciences and the American Physiological Society 17, 56-61
- Shah, D. I., and Singh, M. (2006) Involvement of Rho-kinase in experimental vascular endothelial dysfunction. Molecular and cellular biochemistry 283, 191-199
- Lu, Q. Y., Chen, W., Lu, L., Zheng, Z., and Xu, X. (2014) Involvement of RhoA/ROCK1 signaling pathway in hyperglycemia-induced microvascular endothelial dysfunction in diabetic retinopathy. International journal of clinical and experimental pathology 7, 7268-7277
- 43. Yao, L., Chandra, S., Toque, H. A., Bhatta, A., Rojas, M., Caldwell, R. B., and Caldwell, R. W. (2013) Prevention of diabetes-induced arginase activation and vascular dysfunction by Rho kinase (ROCK) knockout. Cardiovascular research 97, 509-519
- Cicek, F. A., Kandilci, H. B., and Turan, B. (2013) Role of ROCK upregulation in endothelial and smooth muscle vascular functions in diabetic rat aorta. Cardiovascular diabetology 12, 51

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